

# The Role of Tissue-Delivered Dose and Dose Metric for Hazard Grouping and Risk Assessment of Nanomaterials (NMs)

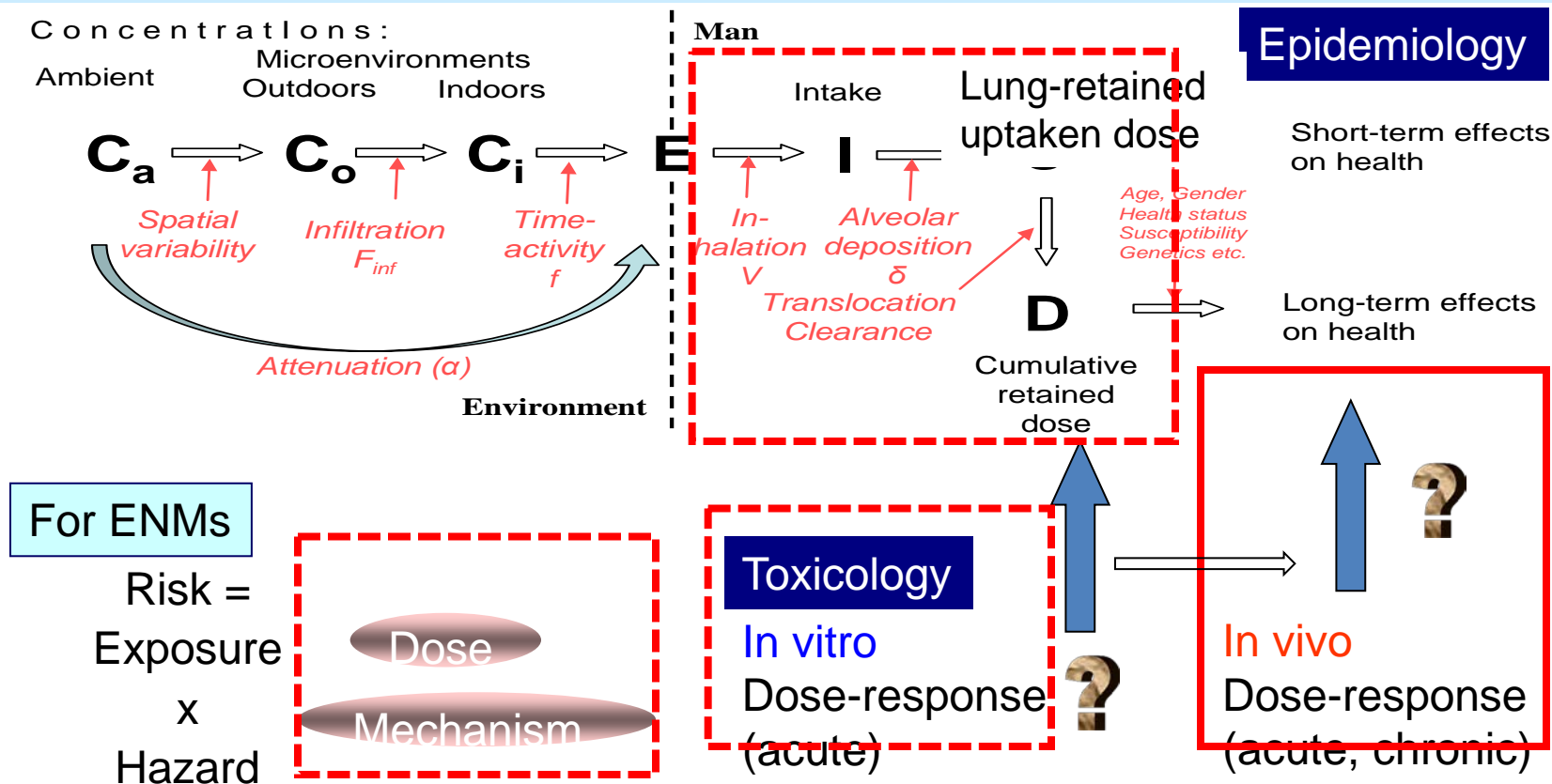
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SmartNanoTox Project Online Conference, 24 June 2020

# From exposure to inhaled NMs to health effects





# Pivotal Role of Tissue Delivered Dose

Schmid, Cassee, Part. Fibre Tox., 14(1):52, 2017

Animal/cell models

Exposure level

Delivered concentration

(Tissue-delivered) Dose - Response Relationship

Lung/Tissue-delivered dose

Toxicological response

Acceptable/desired response or health effect

Maximum allowed tissue-specific dose in model organism (NOAEL/LOAEL)

Animal  $\neq$  Human exposure level

Change in aerosol size distribution & composition

- Coagulation, condensation, evaporation,
- Photoprocessing, gas/medium-particle interaction
- Wall deposition losses, dilution, filtration,

Relating concentration to tissue dose

- Inhaled air/medium volume
- Tissue-deposited aerosol/particle fraction (~size, density, shape)
- Clearance from tissue (~size, shape)

Translation from model to human

- Model-human susceptibility factor
- Safety factor

Acceptable/desired health effect

Human

Exposure limit

Delivered concentration

NOAEL/LOAEL in humans

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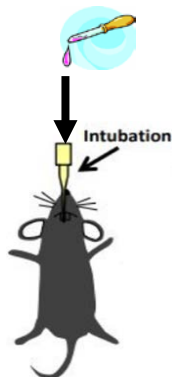
# Tissue-delivered dose

# In vivo – Pulmonary Exposure Technologies and Models

Mouse (C57BL/6)

Intratracheal  
**Instillation**

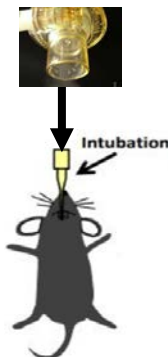
Bulk liquid



Mouse (C57BL/6)

Ventilator-assisted  
**Inhalation**

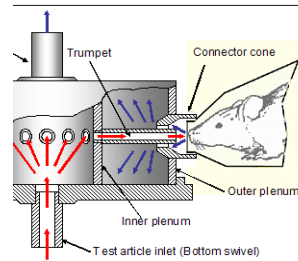
Liquid aerosol



Rat (Wistar)

Nose-only  
**Inhalation**

Liquid/dry aerosol



Dose rate/delivery period  
Substance efficiency  
Degree of physiol. Relevance  
Dosimetry method

High / ~ sec

High / ca. 80%

Low

80% - applied dose

Medium / ~ min

Medium/ ca. 5%

Medium

5% - applied dose  
(account for retainment of  
NM in nebulizer)

Low / ~ hours to months

low / <0.1%

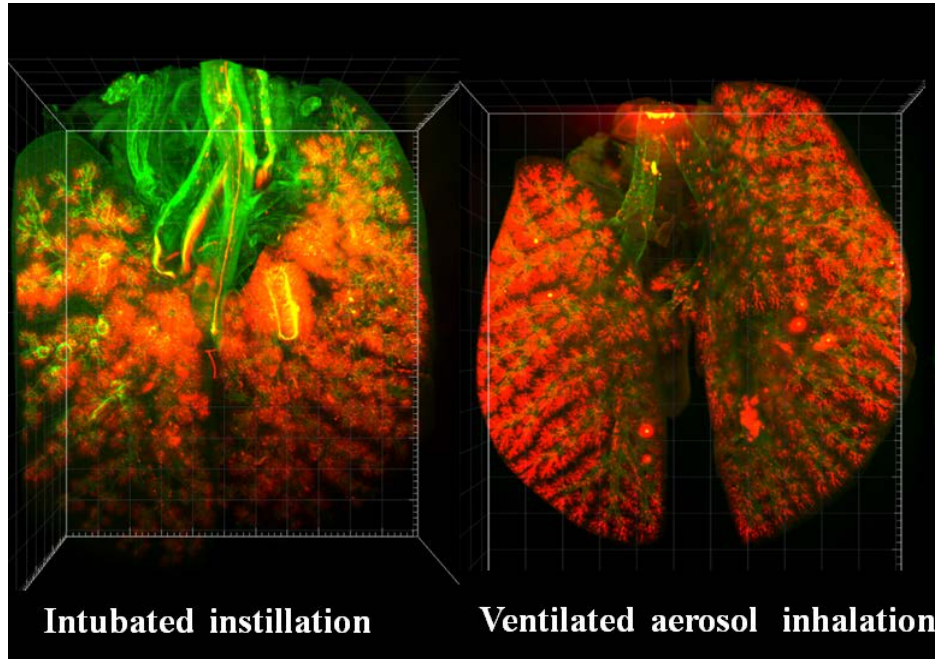
High

- From inhaled conc. & lung deposition model
- ICP-MS, thermogravimetry in lung homogenates

# 3D Co-Mapping of Morphology and Aerosol Deposition in Murine Lung

## 3D Light sheet fluorescence microscopy after optical clearing of tissue

Mouse lung after application of fluorescent nanoparticles

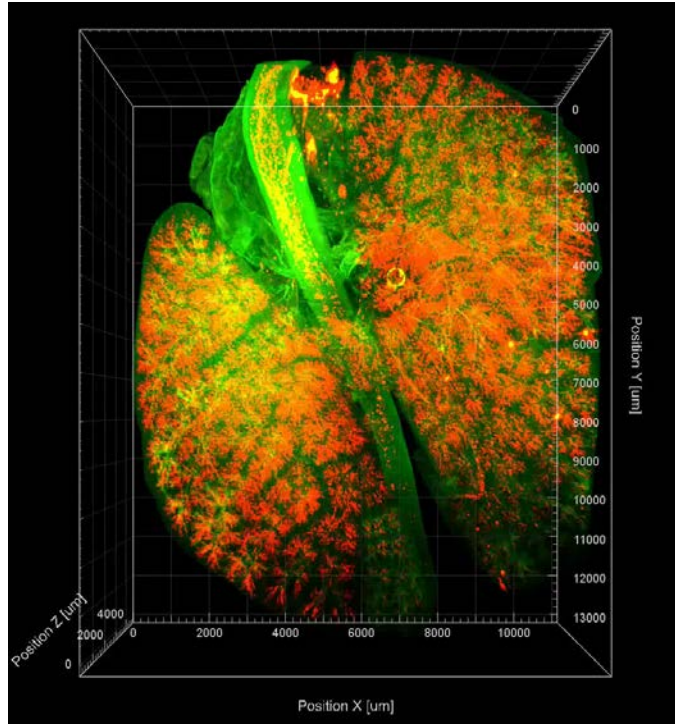


Green – autofluorescence of lung tissue

Red - Fluorescent melamine 600 nm particles

# 3D Co-Mapping of Morphology and Aerosol Deposition in Murine Lung

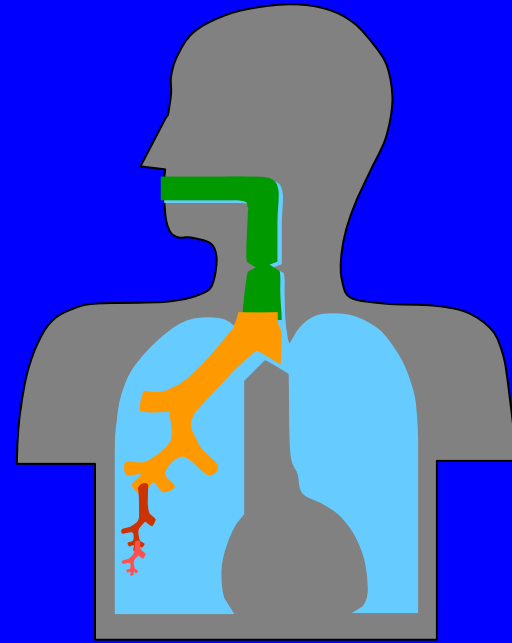
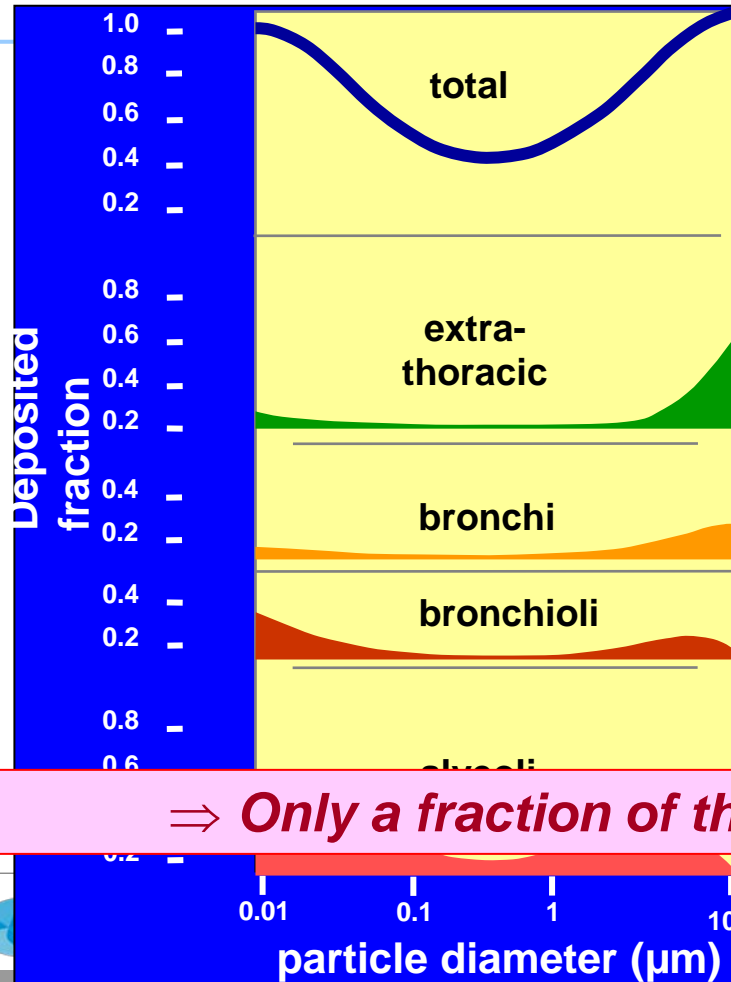
## Ventilator-assisted aerosol inhalation



Green – autofluorescence of lung tissue  
Red - Fluorescent melamine 600 nm particles  
(delivered as suspension in 3 µm droplets)

Yang, ..., Stoeger, Schmid, ACS Nano, 51, 4, 526-535, 2019

# Aerosol Deposition in Human Lung



particle density:  $1 \text{ g cm}^{-3}$

⇒ *Only a fraction of the inhaled dose reaches the cells*



# *In vitro* models: Cells, exposure types, assays

## Pulmonary cell types/assays

**Mouse:** LA4 - epithel cells  
MHS - macrophages  
co-culture: LA4 + MHS

**Rat:** N8383 - macrophages

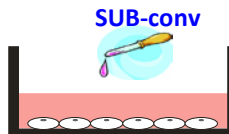
### **Endpoints (24h)**

- Viability (WST-1)

### Cell free Oxidative Potency of NMs

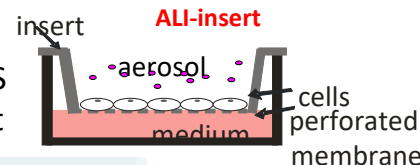
- DCFH assay: Reactive oxygen species (ROS)

## Submerged cell culture



co-culture: LA4 + MHS  
(with whole surfactant)

## Air-liquid interface (ALI)



VitroCell® Cloud



**Exact real-time dosimetry**  
Quartz Crystal Microbalance (QCM)



# *In vitro* particokinetics affects cellular dose

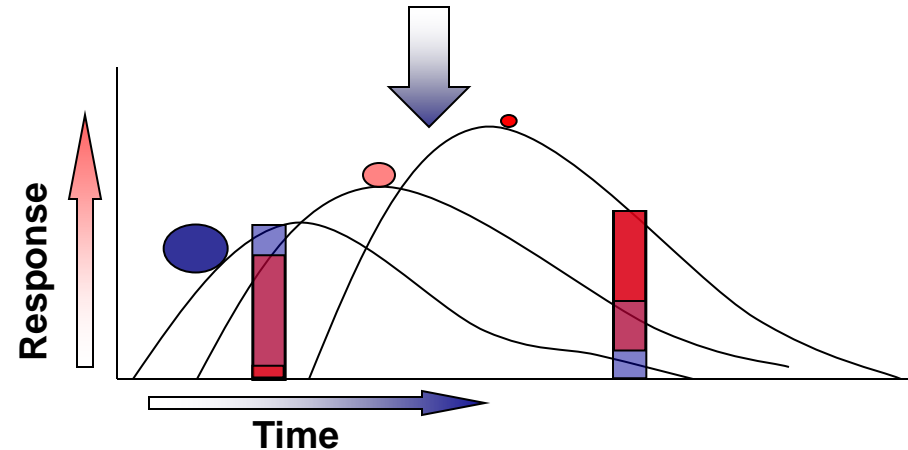
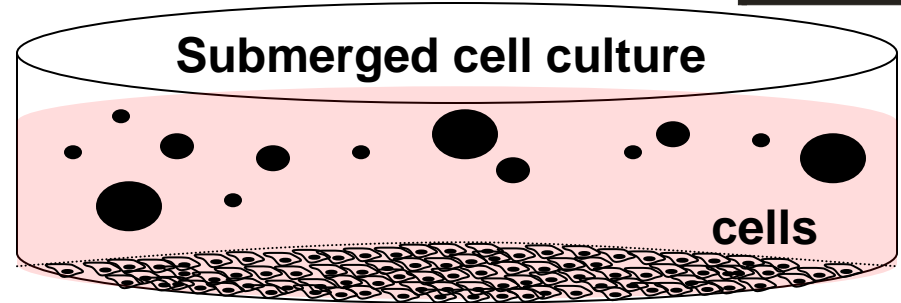
Media "dose" is different than dose to the cell

Shape, size, and density affect

- delivered dose
- delivery rate

Delivery rate/dose impacts timing/magnitude of response

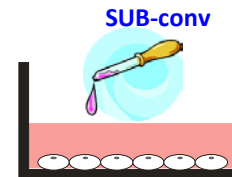
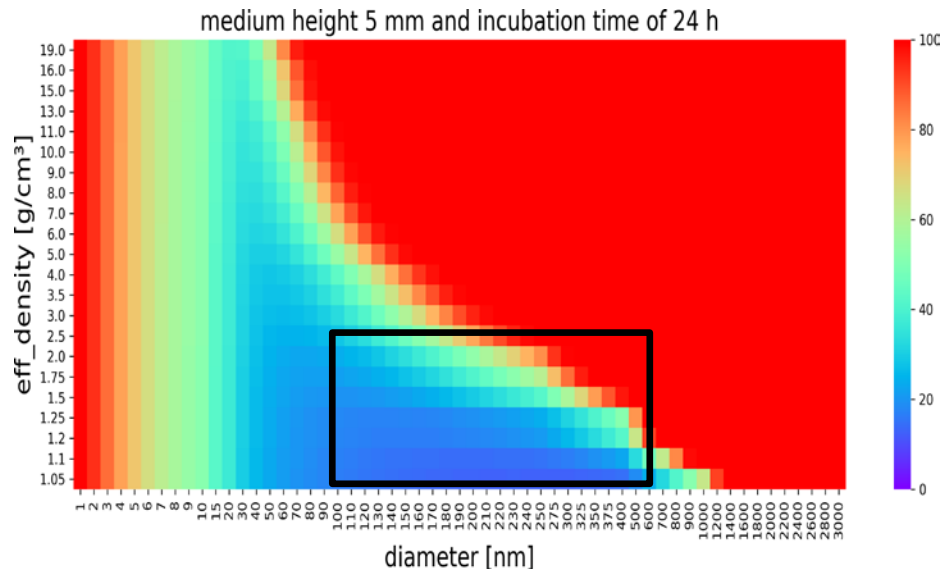
Courtesy J. Teeguarden



# Delivered dose fraction in submersed cell cultures systems

## ISDD particokinetics model

(Hinderliter et al., Part Fibre Tox., 7:36, 2010)



### Typical cell culture conditions:

- Medium height ~ 5 mm
- Incubation time 24h

### Typical NM parameters:

DLS volume diameter: 100 – 600 nm  
Agglomerate density: 1.05 – 2 g/cm<sup>3</sup>

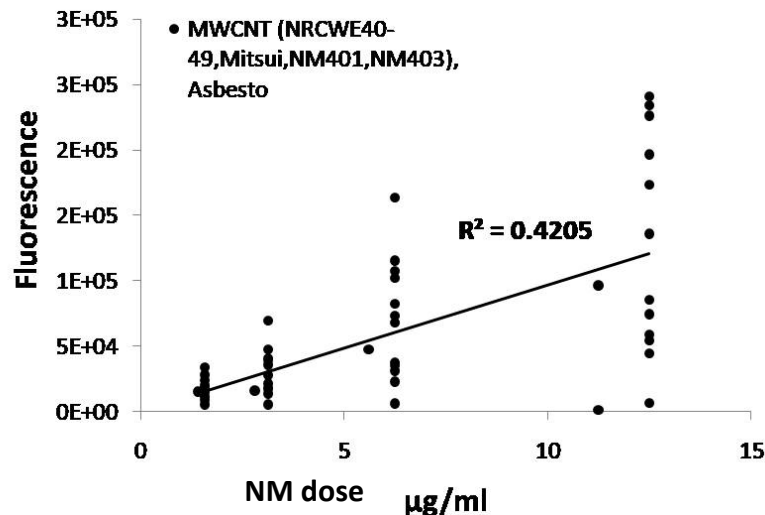
Typical cell-delivered dose fraction:  
<5 – 100%

⇒ *Only a fraction of the nominal dose reaches the cells*

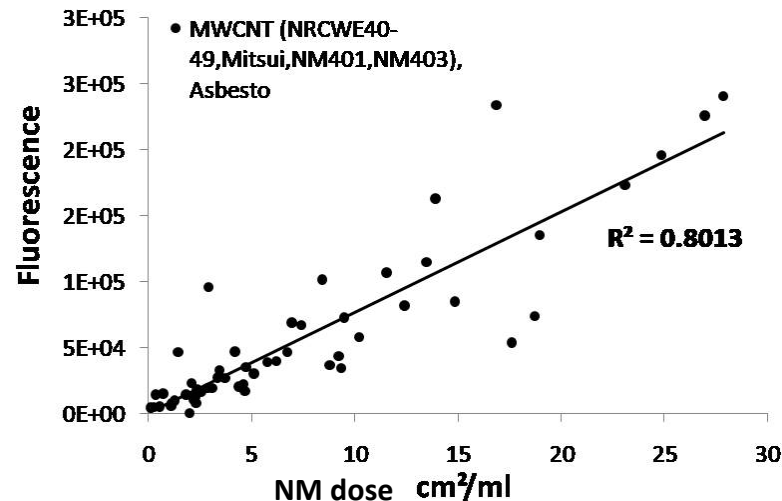
# Dose Metric

# Cell free ROS assay (DCFH) for 13 different fiber-like NMs (12 MWCNTs & asbestos)

## Mass



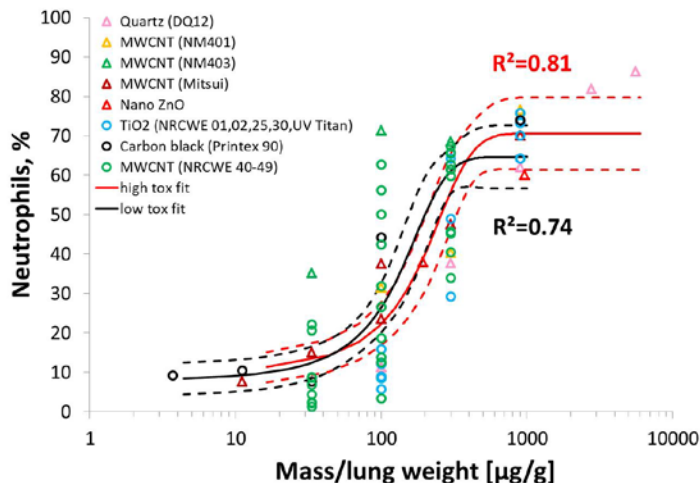
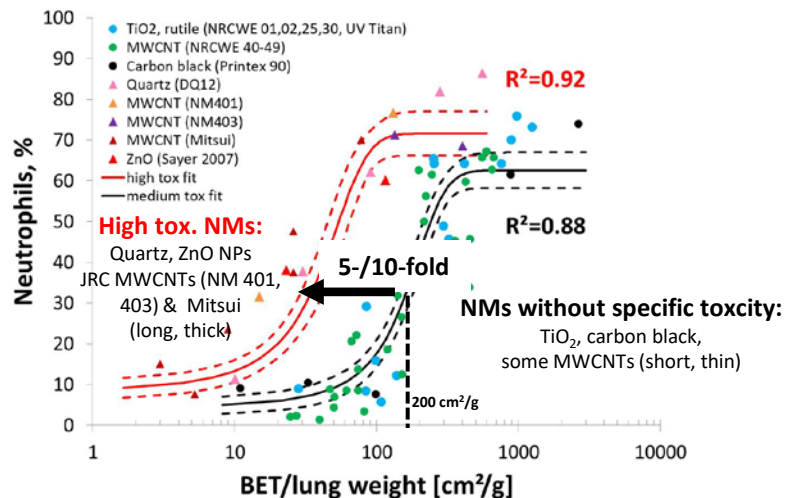
## Surface area



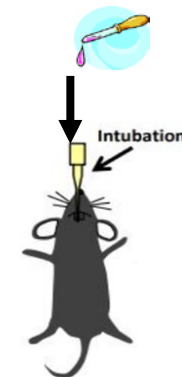
⇒ *Surface area is more predictive of ROS than mass*

# Acute lung inflammation in mice – Instillation of NMs

Number of neutrophils/total cell count in Bronchoalveolar lavage (BAL) 1d after instillation



Bulk liquid

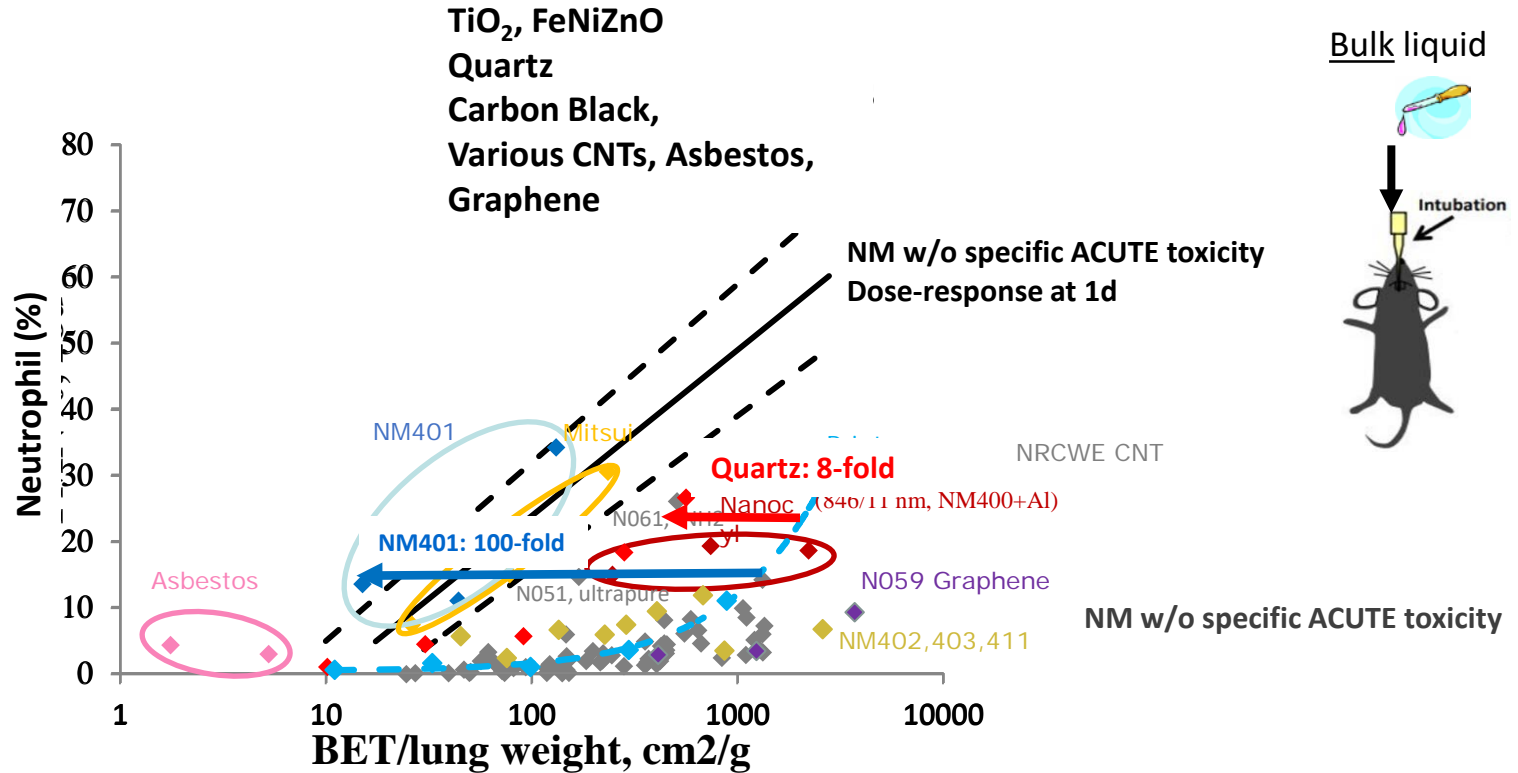


Instillation

⇒ **Surface area: Allows identification of Hazard Classes**

⇒ **Hazard factors (relative to NMs w/o spec. tox): 5 – 10-fold**

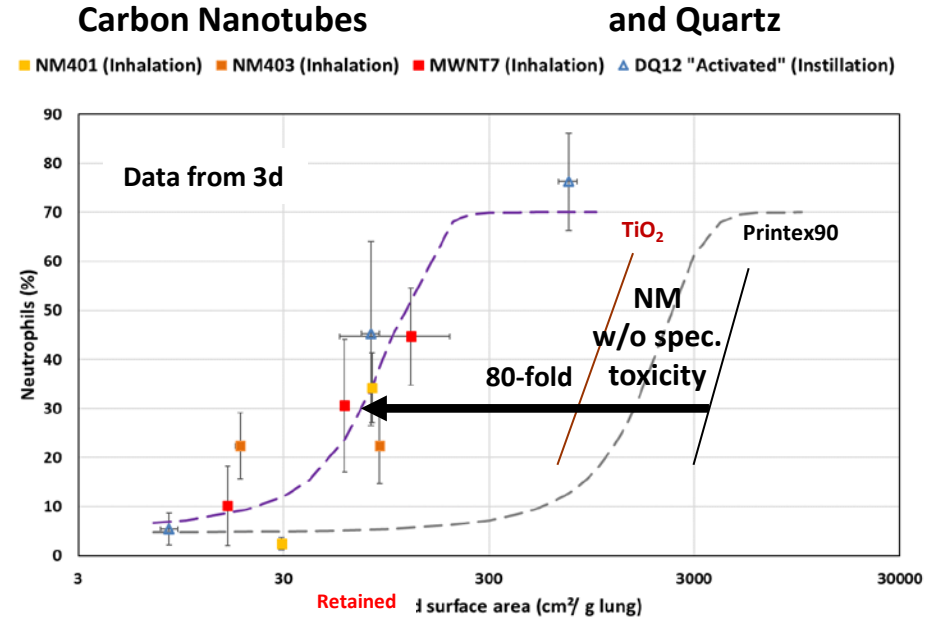
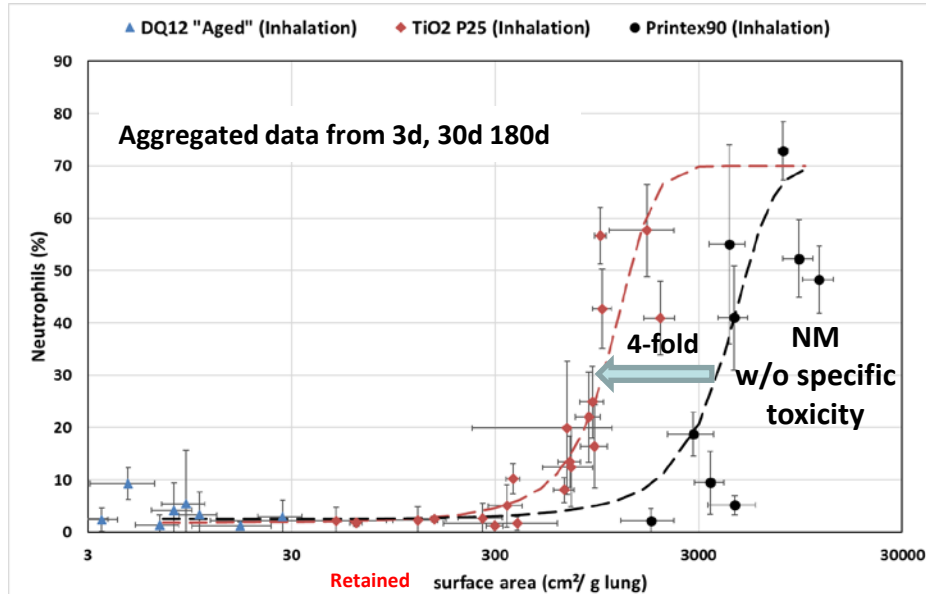
# Sustained inflammation in mice – 28d



⇒ **Similar hazard classes as for acute inflammation (1d)**

# Inflammation after 28d nose-only inhalation (rat)

4 week inhalation (sub-chronic): 6h/d, 5d/w, 4w



⇒ 28d hazard factor after instillation is similar (MWCNT (NM401) or smaller (DQ12) than after nose-only inhalation



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# In vitro / in vivo comparison

# Acute Inflammatory Efficacy (in vivo) - Oxidative potency (in vitro)

In vivo: Mouse

NM instillation

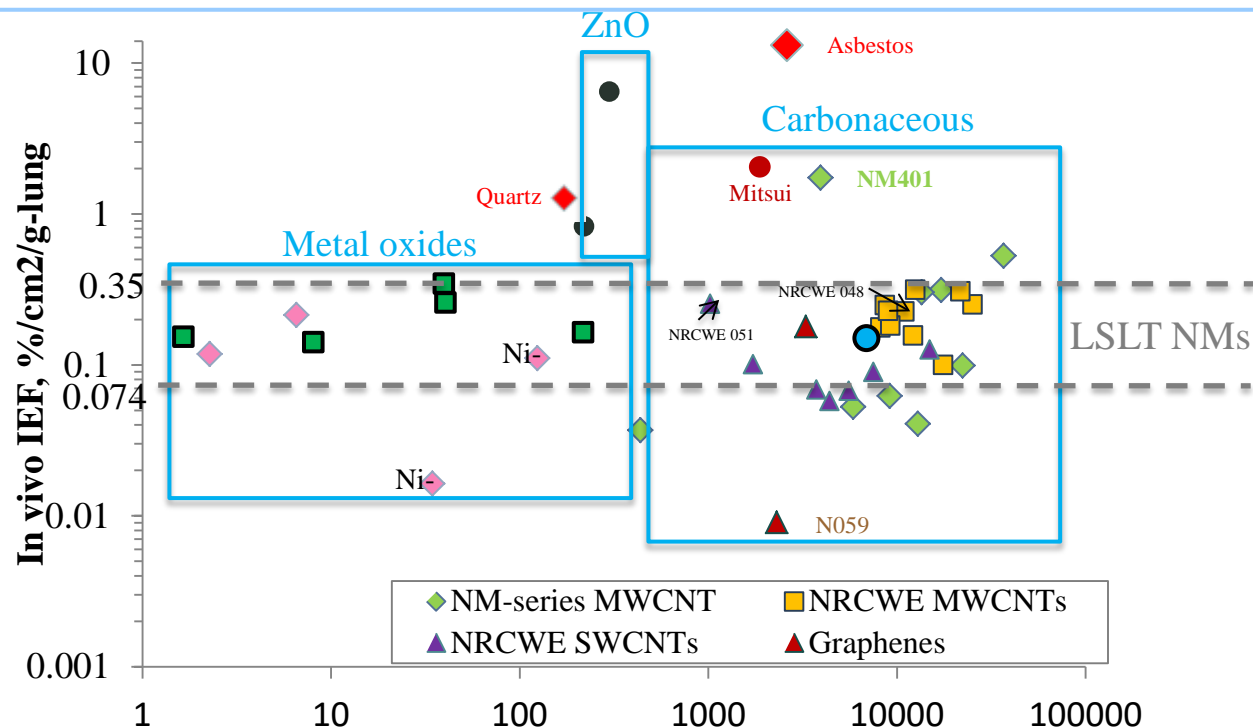
Neutrophil influx (1d)

Slope of Dose-Response

In vitro:

Cell-free ROS (DCFH)

Slope of Dose-Response



⇒ **Cell-free ROS is a good predictor for MetOx vs Carbonaceous NM induced inflammation**

⇒ **Cell-free ROS is NOT a good predictor for acute in vivo inflammation**

**In vitro / in vivo comparison: IC<sub>50</sub> (cm<sup>2</sup>/cm<sup>2</sup>) for *in vitro* viability (WST1, 1d)) and *in vivo* inflammation (neutro.; 1d & 28d)**

Material	BET (m2/g)	Murine (in vitro; submerged)			Rat (in vitro, subm.)	Mouse (in vivo) Instillation (applied dose)		Rat (in vivo) Inhalation
		LA4 (epithel)	MHS (macs)	LA4+ MHS (ALI)		1d	28d	
					N8383 (mac)			28d
Printex 90	272	25	54	>20	400*	0.067	1	1
MWCNT Mitsui-7	26	0.84*	0.38	0.3	63*	0.005	0.05	0.02
MWCNT NM401	18	2.4*	0.25	0.12	44*	0.006	0.03	0.02
ZnO (NM110)	12	0.24	0.18	0.3	0.6*	0.008	---	---
Quartz (DQ12)	10	0.16*	3	>0.9	0.2*	0.01	0.14	0.02

**Printex90:  $f_{\text{haz}} = 1$**

**Low Tox -  $f_{\text{haz}} = 1 - 5$**

**Med. Tox:  $f_{\text{haz}} = 5 - 10$**

**High Tox:  $f_{\text{haz}} = 10 - 100$**

\* IC<sub>50</sub> ~ 4\*IC<sub>12.5</sub>)

⇒ **Mouse instill. (1d) is good qualitative predictor of rat inhalation (28d + 3d)**

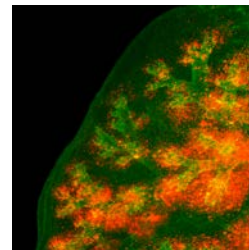
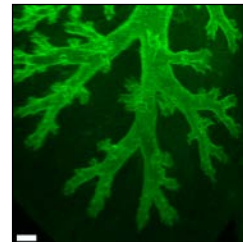
⇒ **Mouse instill. (28d) is good quantitative predictor of rat inhalation (28d + 3d) (3 out of 4)**

⇒ **In vitro cell viability is a good predictor for 28d (+3d) inflammation (MHS; LA4+MHS: 4 out of 4/5)**

⇒ **In vitro cell viability requires higher dose than in vivo inflam. (~ 100-fold (1d) ~ 10-fold (28d))**

# Summary/Conclusions

- ❑ Surface area is a powerful dose metric not only for spherical, but also for fiber-like NMs
  - ❑ Clear classification into NMs with and without specific toxicity
- ❑ Tissue-delivered surface area dose (not exposure concentration) facilitates
  - ❑ Experimental determination of hazard factors
  - ❑ *In vitro* / *in vivo* translation ---- prediction of *in vivo* onset dose from *in vitro* data
- ❑ Instillation (in mice) is a good predictor for inhalation
  - ❑ 1d instillation ~ 1d acute inhalation (ventilator-assisted; in mice)
  - ❑ 28d instillation ~ Subchronic inhalation in rats (28d + 3d)
- ❑ *In vitro* cell viability is powerful predictor of acute (1d) and subchronic *in vivo* inflammation (28d)
- ❑ *In vitro* cell viability requires about 100-fold higher dose than acute inflammation (instillation)
  - ❑ For simple cell culture models (viability assay) we have to accept „non-realistic“ high doses for hazard testing



# Questions/Comments?

